

Essentials of

MEDICAL MICROBIOLOGY

*As per the Competency Based Medical
Education Curriculum (NMC)*

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4th Edition
Revised Reprint



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for TOC & Sample Chapter
on General Bacteriology:
Bacterial Taxonomy



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What is new in this edition?

- Divided into two parts: (1) General Microbiology, Immunology, and Hospital Infection Control; (2) Systemic Microbiology.
- Systemic Microbiology features eight sections with initial chapters on clinical infectious syndromes and detailed chapters on etiological agents.
- Updated Parasitology for Indian Medical Graduates, integrated with respective syndromes.
- Enhanced Hospital Infection Control with new topics like antimicrobial stewardship, PPE protocols, and transmission precautions.
- General Microbiology restructured to include virology, parasitology, mycology, and condensed general bacteriology.
- Overview chapters added for clarity in Systemic Microbiology.
- COVID-19 Chapter-has been revised and updated as per the current scenario.
- Updated AETCOM module for second MBBS students.
- Recent updates on lab diagnosis, treatment guidelines, and vaccines, including tuberculosis and HIV.
- Epidemiology includes recent outbreaks and geographical distribution with a focus on Indian epidemiology.
- Latest references from textbooks and health guidelines.
- Increased use of tables, flowcharts, images, and diagrams.
- Designed for MBBS and NExT exams with a concise, bulleted format.
- Simple language with summary boxes for lab diagnosis and treatment; includes clinical case-based essays and MCQs.

General Bacteriology: Bacterial Genetics

CHAPTER 3.4

CHAPTER PREVIEW

- Principles of Bacterial Genetics
 - Bacterial DNA
 - Bacterial RNA
 - Polypeptide Synthesis

- Plasmid
- Mutation
- Horizontal Gene Transfer
 - Transformation
 - Transduction

- Lysogenic Conversion
- Conjugation
- Transposition
- Gene Transfer by Artificial Methods

Chapter Previews are given at the beginning of each chapter

Feature Tables to simplify complex topics in structured parts

Table 3.3.1: Types of infections and various specimens collected.

Type of infections	Specimens collected
Bloodstream infection, sepsis, endocarditis	Paired blood culture specimens <ul style="list-style-type: none"> Collected aseptically by two-step disinfection of skin; first with alcohol followed by chlorhexidine 8–10 mL of blood (for adults) collected in blood culture bottles
Infectious diseases requiring serology	Blood (2 mL/investigation) <ul style="list-style-type: none"> Collected by minimal asepsis (one-step skin disinfection with alcohol) Collected in vacutainer
Diarrheal diseases	Stool (mucus flakes), rectal swab
Meningitis	Cerebrospinal fluid (CSF)
Infections of other sterile body area	Sterile body fluids; e.g. pleural fluid, synovial fluid, peritoneal fluid
Skin and soft tissue infections	Pus or exudate, wound swabs, aspirates from abscess and tissue bits
Anaerobic infections	Aspirate, tissue specimens, blood and sterile body fluids, bone marrow (swabs, sputum not satisfactory)
Upper respiratory tract infections	Throat swab with membrane over the tonsil, nasopharyngeal swab, per-nasal swab
Lower respiratory tract infections	Sputum, endotracheal aspirate, bronchoalveolar lavage (BAL), protected specimen brush (PSB) and lung biopsy
Pulmonary tuberculosis	Sputum—early morning and spot <ul style="list-style-type: none"> Collected in well-ventilated area Gastric aspirate for infants
Urinary tract infections	Midstream urine <ul style="list-style-type: none"> Suprapubic aspirated urine Catheterized patient—collected from the catheter tube, after clamping distally and disinfecting; not from urubag
Genital infections	Urethral swab, cervical swab—for urethritis <ul style="list-style-type: none"> Exudate from genital ulcers
Eye infections	Conjunctival swabs <ul style="list-style-type: none"> Corneal scrapings Aqueous or vitreous fluid
Ear infections	Swabs from outer ear <ul style="list-style-type: none"> Aspirate from inner ear

Bacteriology: Laboratory Diagnosis of Bacterial Infections

Specimen Storage before Processing

Most specimens can be stored at room temperature immediately after receipt, for up to 24 hours. However, there are some exceptions.

- Blood cultures**—should be incubated at 37°C immediately upon receipt
- Sterile body fluids**, bone, vitreous fluid, synovial fluid, peritoneal fluid—should be immediately placed upon receipt and stored at 37°C
- Corneal scrapings**—should be immediately placed at bedside on a blood agar and chocolate agar
- Stool culture**—can be stored up to 72 hours at 4°C
- Urine** (mid-stream and from the catheter), **lower respiratory tract specimens**, **gastric biopsy** (for *Helicobacter pylori*)—can be stored up to 24 hours at 4°C.

DIRECT DETECTION

Direct detection of bacteria in the clinical specimen plays a very important role in early institution of antimicrobial therapy. These methods include microscopic demonstration of bacteria—using wet mounts and other methods such as detection of antigens or nucleic acid in the clinical specimen.

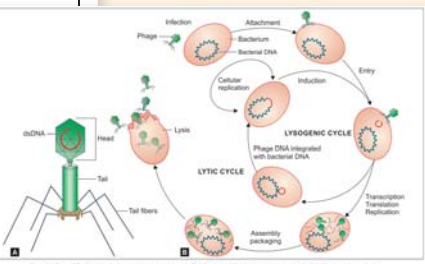
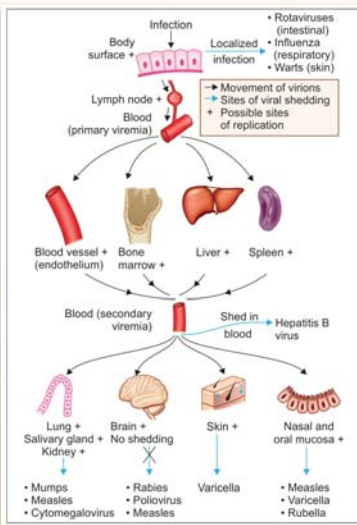
STAINING TECHNIQUES

Structural details of bacteria cannot be seen under a light microscope due to lack of contrast. Hence, it is necessary to use staining methods to produce color contrast and thereby improve the visibility. Before staining, the smears are fixed so that they will not be displaced during the staining process. Fixation also protects the internal structures of cells in a fixed position. It is done by two methods.

- Heat fixation** is done by gently flame heating an air-dried film used for bacterial smears.
- Chemical fixation** (used for fixed smears).

Common staining techniques used in diagnostic bacteriology include:

- Simple stain** Basic dyes, such as methylene blue or basic fuchsin are used as simple stains. They provide the color contrast, but impart the same color to all the bacteria in a smear.
- Negative staining** A drop of bacterial suspension is mixed with dyes, such as India ink or nigrosin. The background gets stained black whereas unstained bacterial yeast capsule stand out in contrast. This is very useful in the demonstration of bacterial yeast capsules which do not take up simple stains.
- Immunofluorescence** Bacterial cells and structures that are too thin to be seen under the light microscope, are thickened by impregnation of silver salts on their surface to make them visible, e.g. for demonstration of bacterial flagella and spores.



Illustrative Diagrams & Figures are given for better understanding

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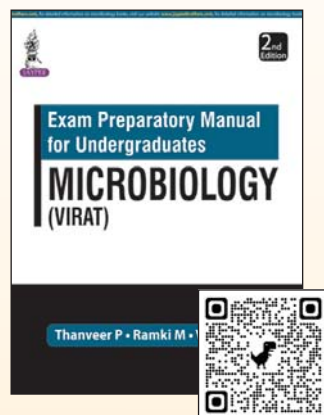
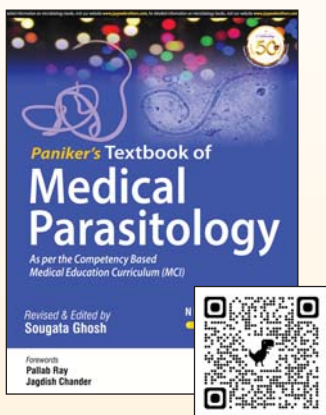
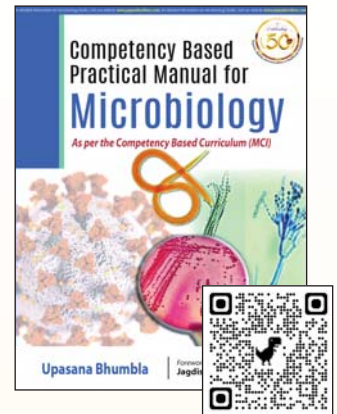
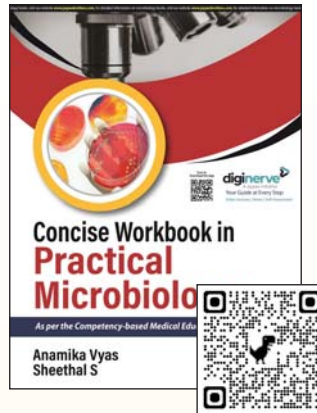
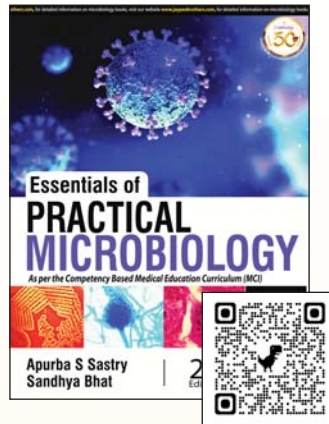
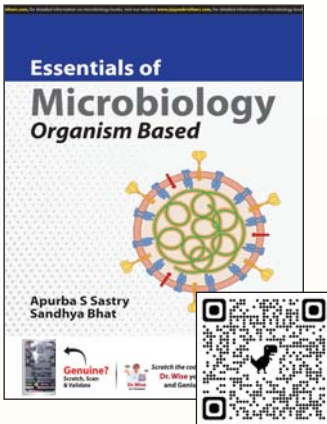
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